ABSTRACT
Normoblasts may be seen in peripheral blood smear of newborns. The number of normoblasts per 100 white blood cells is generally in the range of 0-10. They can be seen more common than usual in hypoxic condition, because intrauterine hypoxia increases the production of red blood cells. However, multinucleated normoblasts in a newborn caused by hypoxia haven’t been reported before. We present a newborn with multinucleated normoblasts secondary to intrauterine hypoxia. This case is important; because it is the first time multinucleated normoblasts in peripheral blood smear of a hypoxic newborn has been detected.

**Key words:** multinucleated normoblasts, hypoxia, newborn.

INTRODUCTION
Normoblast is a precursor in erythroid series. These nucleated red blood cells can be seen in peripheral blood smear of newborns. The number of normoblasts per 100 white blood cells is generally in the range of 0-10. If there is a hypoxic condition, normoblast count may be higher than 10, because hypoxia is the major regulator of production of red blood cells (RBCs).

Multinucleated normoblasts have been demonstrated in bone narrow aspiration smears of patients with congenital dyserythropoietic anemia (CDA) type II and type III; in peripheral blood smear of a patient with CDA type II. Multinucleated normoblasts can also be seen in myelodysplasia and erythroleukemia. However, hypoxia hasn’t been presented as a reason of multinucleated normoblasts until now. Here, we report this case of multinucleated normoblastemia of a newborn secondary to hypoxia.

CASE REPORT
Following an uneventful pregnancy, a full-term (38 weeks), male infant was born by spontaneous vaginal delivery to a 38-year-old gravida 4, para 4 mother. Umbilical cord had wrapped around his neck on the delivery. Apgar scores were 6 and 9 at 1 and 5 minutes. Amniotic fluid was clear. Birth weight was 3900 g. The baby presented with tachypnea and poor sucking at 6th hour of his life. Blood sample was taken. In automated complete blood count, his hemoglobin (Hb) was 19 g/dL; white blood cell count, 96.5 x109/L; and platelets count, 55x109/L. The baby was referred from a local hospital because of probable leukemia and was admitted to the neonatal intensive care unit of our hospital.

On admission, the vital signs were normal. Physical examination revealed dry skin, widespread desquamation, decreased subcutaneous fat, subconjuctival hemorrhage, and liver 2-cm below right costal margin. His complete blood count analysis showed: Hb 18.5 g/dL; white blood cell count, 79x109/L; and platelets count, 50x109/L. In his peripheral blood smear; the number of normoblasts was 360 per 100 white blood cells. Most of these normoblasts were binucleated or multinucleated (Figure 1). The real leukocyte count was approximately 17x109/L; the presence of numerous normoblasts have increased the white blood cell count in automated complete blood count analyzer. The biochemical markers were: total bilirubin/direct bilirubin was 13.1/0.7mg/dL; blood urea nitrogen, 13.8 mg/dL, and creatinine, 1.7 mg/dL. Venous blood gas analysis revealed that pH was 7.24; pCO2, 34 mmHg; pO2, 40 mmHg; and HCO3, 17 mmol/L.
There wasn’t any ABO incompatibility and direct Coombs test was negative.

Oral feeding and intravenous hydration were administered to the baby. On the 2nd day of hospitalization the baby received phototherapy when total bilirubin level was 15.6 mg/dL. On the 5th day of hospitalization, blood urea nitrogen and creatinine levels became to normal values. Blood urea nitrogen decreased to 3.1 mg/dL and creatinine decreased to 0.59 mg/dL. Then, the intravenous hydration support was stopped. Abdominal and cranial ultrasounds were performed by a radiologist and evaluated as normal. Baby’s activity and sucking became much better day-by-day. On the 10th day of hospitalization, the baby was discharged. There was no information about his follow-up.

**DISCUSSION**

Normoblasts can be seen in a range of 0-10 for every 100 WBC in peripheral blood smear in newborns during the first 5 days of their life.1 Due to the immaturity of reticuloendothelial system, hyposplenism may occur in neonatal period. Generally, normoblasts in circulation, which leak from bone marrow, are cleared in spleen. That’s the reason why normoblasts can be seen in peripheral blood smears of newborns.1

| Figure 1. Multinucleated normoblasts in peripheral blood smear |

**Table 1. Normoblast and WBC count in automated complete blood count analyzer**

<table>
<thead>
<tr>
<th></th>
<th>1st day</th>
<th>2nd day</th>
<th>4th day</th>
<th>5th day</th>
<th>10th day</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normoblast count (per 100 WBC)</td>
<td>360</td>
<td>320</td>
<td>340</td>
<td>280</td>
<td>2</td>
</tr>
<tr>
<td>WBC count in automated complete blood count analyzer (x10^9/L)</td>
<td>79.3</td>
<td>54.4</td>
<td>42.7</td>
<td>26.9</td>
<td>12</td>
</tr>
</tbody>
</table>

Furthermore, if cellular components increase, the capacity of spleen is overcomed and normoblasts can be seen more than usual in peripheral blood smears.1 In conditions with an increase in production of erythroid precursors including maternal diabetes mellitus, intrauterine growth retardation, and hypoxia; normoblast count can be more than 10 for every 100 WBC.1,3

At hypoxia, depending on the decrease of oxygen content in tissues, erythropoietin increases for red blood cell production. Therefore; high erythropoietin levels result in elevated production of erythroblasts in bone narrow and there can be more normoblasts in peripheral blood smear.6

Previous studies showed that normoblasts could be a marker of fetal hypoxia.2,3,7,8 According to the study reported by Fox H et al., there can be a relationship with normoblast count and duration of hypoxia.7 In case of a short duration of hypoxia, there can not be enough time to increase normoblast production.7 The time between the increase of erythropoietin caused by hypoxia and the appearance of normoblasts should be at least 36-48 hours.9 Therefore, in our case, very high levels of normoblasts may be explained not only by acute hypoxia due to wrapping of the cord, but also by intrauterine chronic hypoxia. The findings of dysmaturity also supported this idea, because intrauterine hypoxia is known a cause of dysmaturity.4

The distinctive feature of this case is the presence of multinucleated normoblasts in peripheral blood smear. Multinucleated normoblasts have also been demonstrated in patients with congenital dyserythropoetic anemia tip II and type III.10 Normocytic mild anemia, jaundice, splenomegaly, or hepatomegaly is typically seen in patients with CDA type II.5 Liljeholm et al., have shown failure of cytokinesis occurs due to KIF23 gene mutation in CDA type III,11 and KIF23 gene encodes a protein that regulates cell separation during mitosis.5 Therefore multinucleated normoblasts can be seen in patients with CDA type III.11 Diagnoses of CDA type II and III are generally made later in life, because their symptoms are milder. Besides,
erythroleukemia and myelodysplasia maybe other reasons of multinucleated normoblasts. However, our case could not be explained by these conditions. If dyserythropoiesis was the reason for multinucleated normoblasts, decreased hemoglobin level and anemia would be seen. However, hemoglobin levels of our patient were always in a normal range. Moreover, the patient’s symptoms that began in neonatal period were transient; which was not suggestive for CDA type II or III. The reason of multinuclearity may be the increase in production of erythroblasts secondary to hypoxia. For the first time, multinucleated normoblasts have been shown secondary to hypoxia. It seems reasonable to assume that chronic hypoxia and additional acute hypoxia due to cordon wrapping may be the cause for multinucleated normoblasts seen in our patient’s peripheral blood smears.

REFERENCES